

# **TFPoppy**

a.k.a HPAR-3OM

Reference 555670-250 Quantity 250 nmol

Store at 2-8 °C

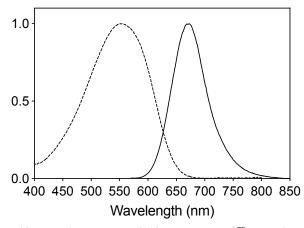
# Properties of TFPoppy when bound to frFAST

Excitation wavelength	555 nm
Emission wavelength	670 nm
Molar absorption coefficient	$45,000  \mathrm{M}^{-1} \mathrm{cm}^{-1}$
Fluorescence quantum yield	21 %
Affinity constant at 25° C	1 μΜ

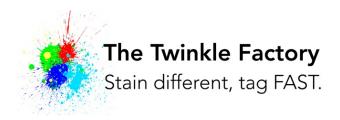
<sup>TF</sup>Poppy is a membrane-permeant fluorogenic ligand that can be used to selectively label frFAST-tagged proteins in solution, in living cells and in fixed cells. <sup>TF</sup>Poppy is almost non-fluorescent when free in solution, but strongly fluoresces when bound to frFAST. <sup>TF</sup>Poppy was designed together with a variant of FAST, frFAST, specifically for far-red labeling. It exclusively works with this variant and should not be used with FAST1 nor FAST2 nor pFAST. This package includes 250 nmol of <sup>TF</sup>Poppy, enabling to prepare 50 mL of a 5 μM labeling solution.

The Twinkle Factory labeling technology is a novel tool that enables the specific fluorescent labeling of any protein of interest. This technology is based on the instantaneous formation of a fluorescent molecular assembly between the small (14 kDa) protein tag FAST and various fluorogenic ligands (TFFluorogens). TFFluorogens strongly fluoresce only when bound to FAST, enabling to detect and image FAST-tagged proteins with high contrast without the need of washing the excess of fluorogenic ligands. The labeling of FAST-tagged proteins with a TFFluorogen is non-covalent and can be reversed if necessary by washing.

The use of the Twinkle Factory labeling technology implies cloning and expressing of the FAST-tagged protein, and labeling the resulting fusion with the <sup>TF</sup>Fluorogen of choice. The labeling of FAST-tagged proteins is described below. Cells expressing FAST-tagged proteins are not supplied by The Twinkle Factory. Note that proteins of interest can be expressed with FAST as either an N- or a C-terminal fusion.



Absorbance (dotted line) and emission (solid line) spectra of TFP oppy bound to frFAST



# Protocol of labeling in living cells

Dissolve one vial of <sup>TF</sup>Poppy in 50  $\mu$ L of DMSO to yield a 5 mM stock solution. Mix by vortexing for few seconds until all the <sup>TF</sup>Poppy is dissolved. Note that different stock concentrations can be made depending on your requirements. <sup>TF</sup>Poppy is soluble in DMSO up to at least 50 mM.

Dilute the stock solution 1:500 in medium or buffer to yield a 10  $\mu$ M labeling solution. Mix thoroughly by vortexing. For best performance, add <sup>TF</sup>Poppy to serum-free medium or buffer, and do not keep/store the labeling solution. Note that different concentrations can be made depending on your requirements. Optimal concentrations range from 1 to 10  $\mu$ M.

Remove the cell culture medium, wash with D-PBS, and replace the buffer with the labeling solution. Incubate for 15-30 seconds and image the cells directly.

Image the cells using appropriate settings. frFAST-tagged proteins labeled with <sup>TF</sup>Poppy have an excitation maximum at 555 nm and an emission maximum at 670 nm.

To reverse the labeling, remove the labeling solution, wash with D-PBS, and replace with culture medium.

# Protocol for labeling in fixed cells

Cells expressing frFAST-tagged proteins can be fixed before labeling with standard fixation methods such as paraformaldehyde, ethanol, methanol. Once the fixation is performed, wash cells with D-PBS, and replace the buffer with a labeling solution (prepared in D-PBS). Incubate for 15-30 seconds and image the cells directly as above. To reverse the labeling, remove the labeling solution and wash with D-PBS.

#### Storage

Dry <sup>TF</sup>Poppy should be stored at 2-8 °C in the dark. Once dissolved in DMSO, the solution should be aliquoted to avoid repeated freeze/thaw cycles and stored at – 20 °C in the dark. With proper storage, <sup>TF</sup>Poppy should be stable at least three years dry or 6 months dissolved in DMSO.

### **Purity and Characterization**

Purity of  $^{\text{TF}}$ Poppy was determined to be > 99% by nuclear magnetic resonance (NMR) and elementary analysis.

#### References

Angew. Chem. Int. Ed. 59, 17917-17923 (2020)

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- EP 3 164 411; JP 2017-527,261; US 10,138,278 (Fluorogen activating and shifting tag (FAST))
- EP 3 719 007; US 2022-0169682 (Split photoactive yellow protein complementation system and uses thereof)

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